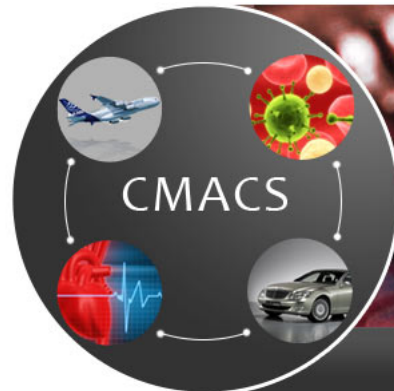


Virtual PI Meeting

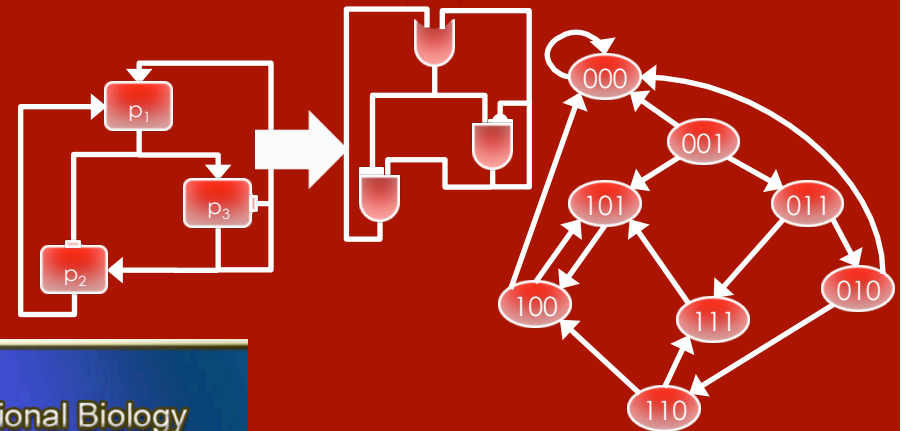


Computational Modeling and Analysis for Complex Systems

April 2012



Department of Computational Biology



Timing Matters in T Cell Differentiation

Natasa Miskov-Zivanov
University of Pittsburgh

Acknowledgements



- **Faeder Lab:**

- Department of Computational and Systems Biology, School of Medicine, University of Pittsburgh
 - John Sekar, James Faeder

- **Morel Lab:**

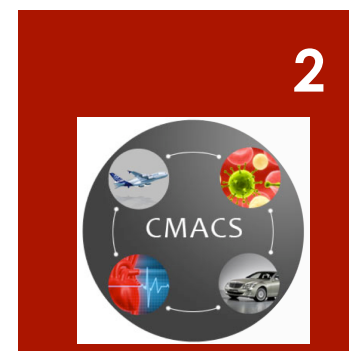
- Department of Immunology, School of Medicine, University of Pittsburgh
 - Michael Turner, Penelope Morel

- **Clarke Lab:**

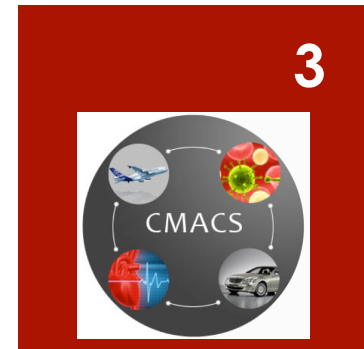
- Computer Science Department, Carnegie Mellon University
 - Paolo Zuliani, Haijun Gong, Edmund Clarke
 - Deepa Sathaye, Alexander Moser

- **Funding:**

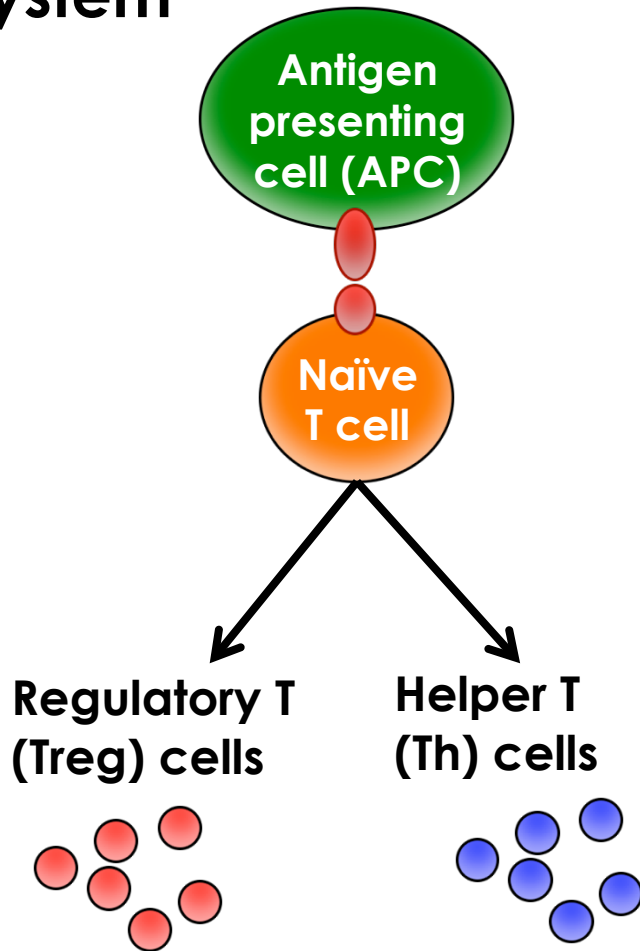
- NSF (Expeditions in Computing)
- NIH



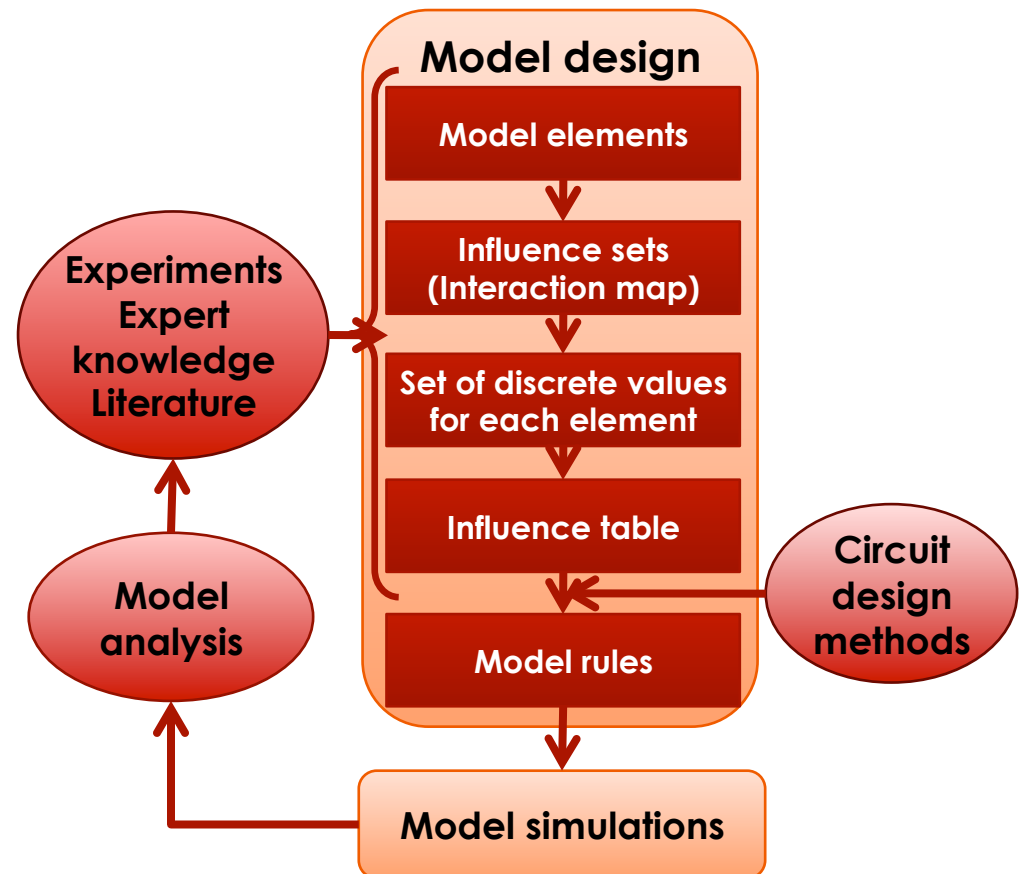
Outline



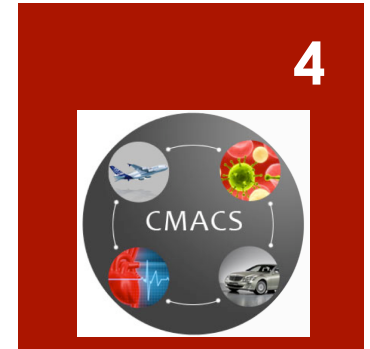
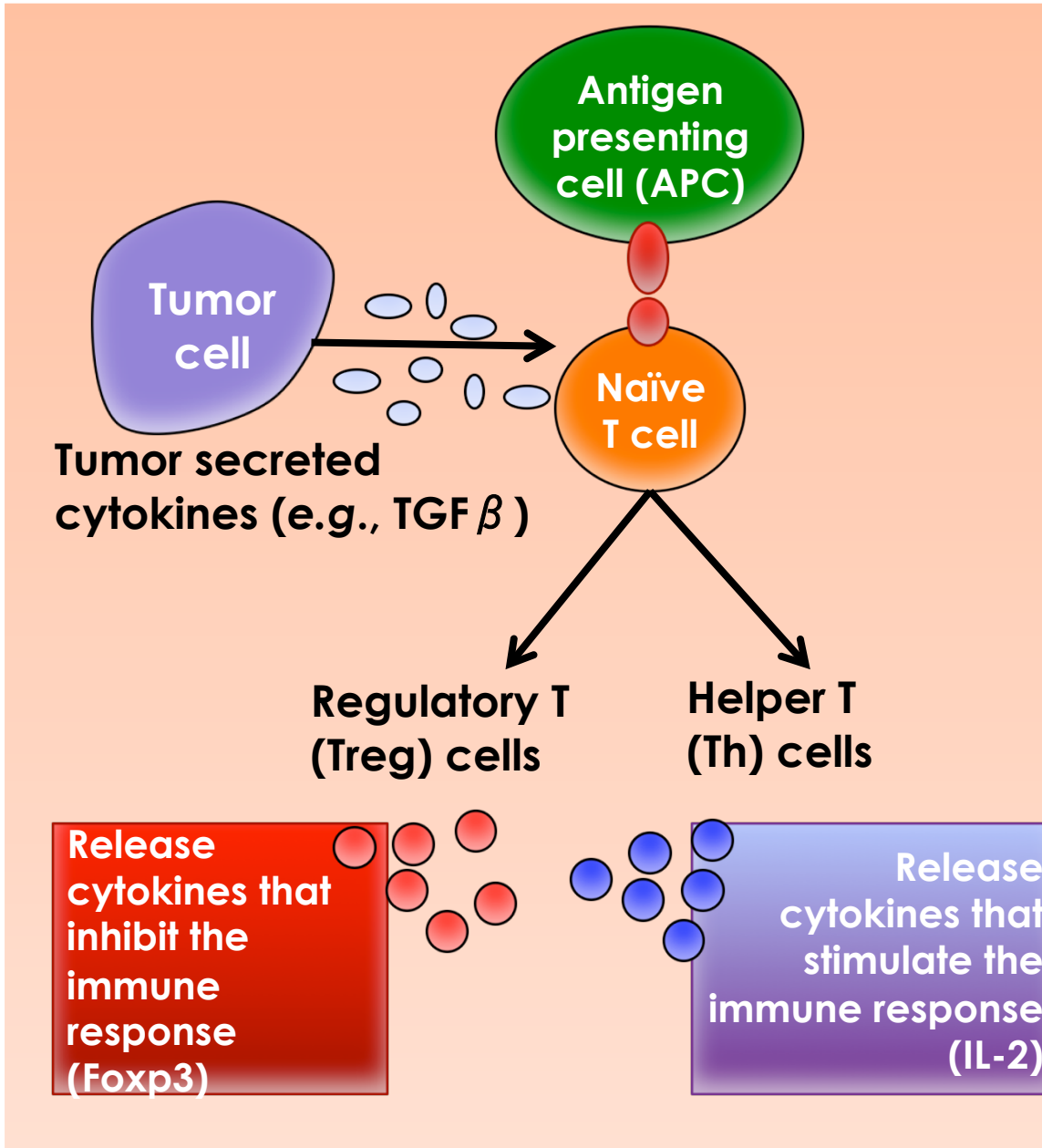
System



Methodology

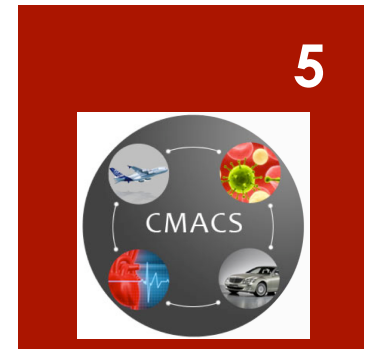


T cell differentiation



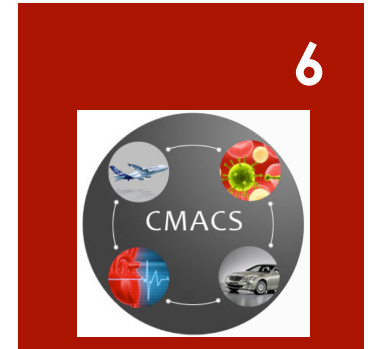
- T cell subpopulation ratios are critical for numerous immune and auto-immune pathologies
- Determinants of the peripheral T cell differentiation are not completely understood

Questions to address



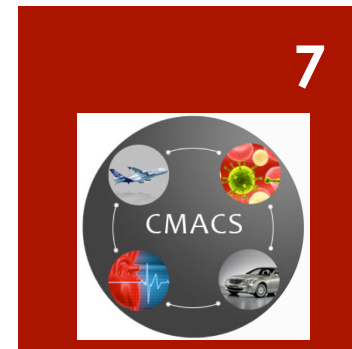
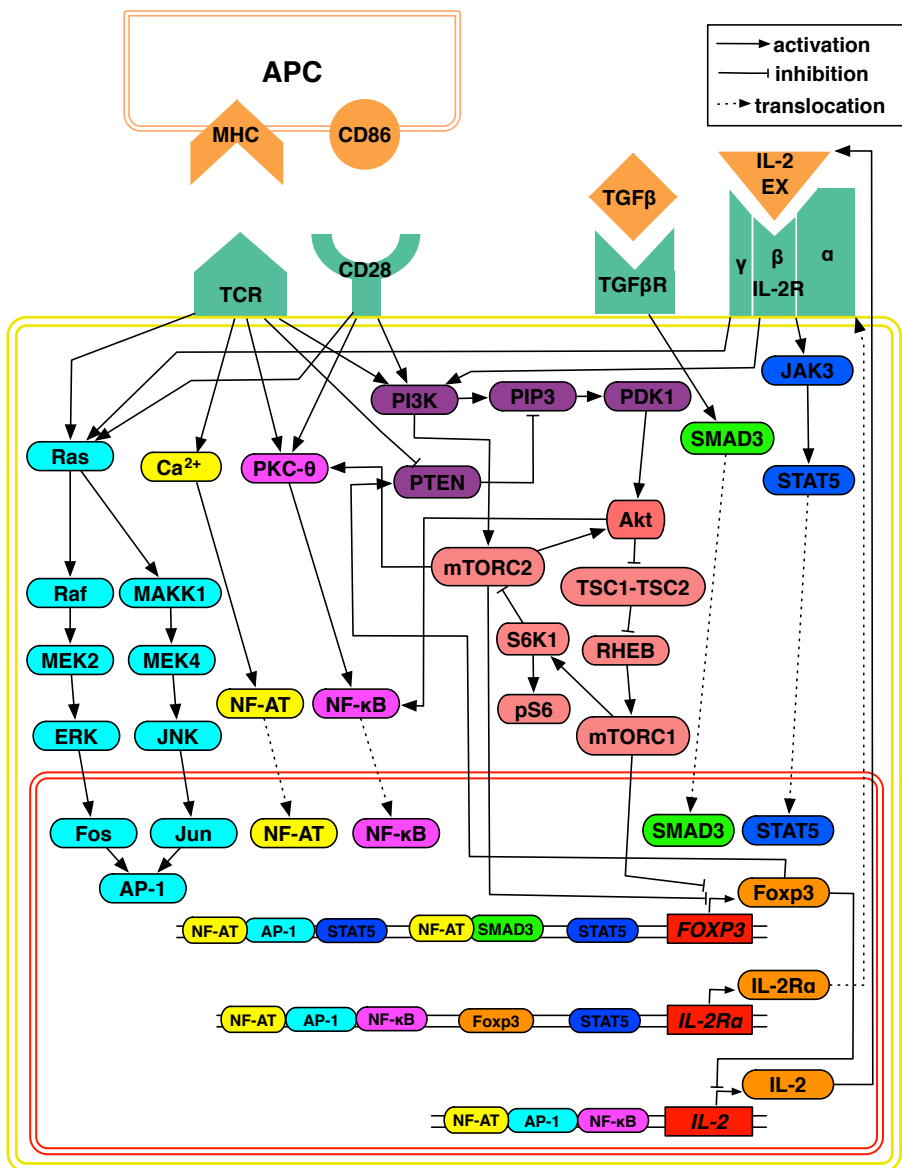
- How can we predict the antigen dose that will induce Treg versus Th?
- Are there signaling cascades in T cells that are critical in this cell fate decision?
- Can we use modeling to identify the critical factors?
- Many clinical trials involving DC vaccines do not take antigen dose into account
- Could also be important for the *ex vivo* expansion of Treg for therapeutic purposes

Modeling goals



- Determine whether known mechanisms are *sufficient* to explain experimental observations
- Suggest *additional experiments* to identify missing mechanisms and clarify areas of *uncertainty*
- Identify *early markers* of the response

Network model



Receptors:

- T cell receptor (TCR)
- Co-stimulation through CD28
- IL-2 receptor (IL-2R)
- TGFβ receptor (TGFβ R)

Transcription factors:

- AP-1, NFAT, NFκB, SMAD3, STAT5

Genes:

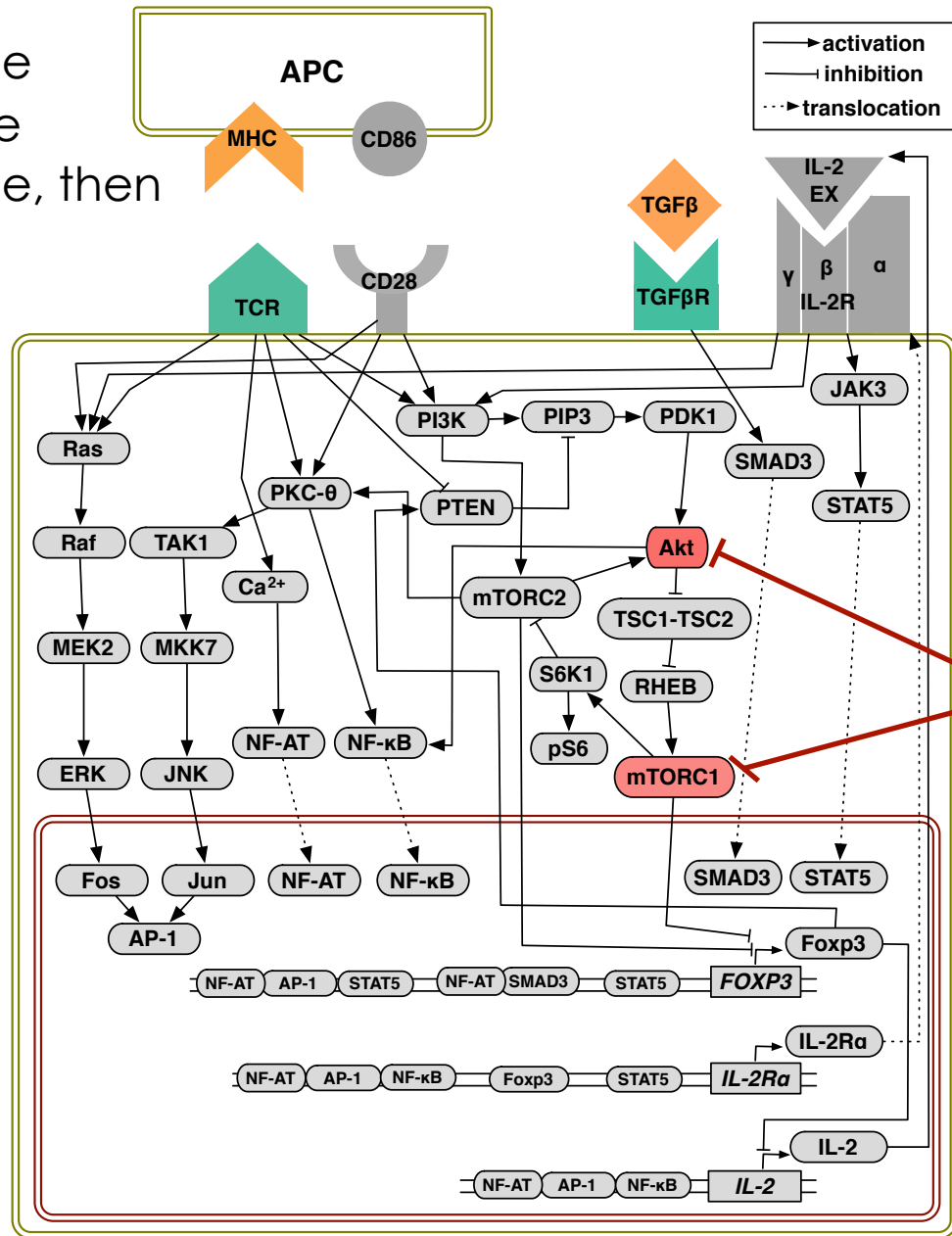
- IL-2, CD25, Foxp3

Other important elements:

- PTEN, PI3K, PIP3, PDK1,
- Akt, mTORC1, mTORC2, TSC1-TSC2, Rheb, S6K1, pS6

Five scenarios

1. High antigen dose
2. Low antigen dose
3. High antigen dose, then removed

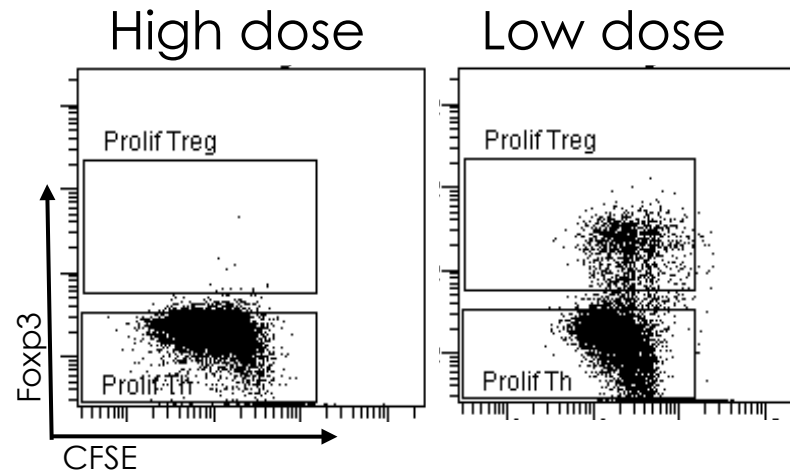


4. High antigen dose and TGFβ

5. High antigen dose, then inhibitors added

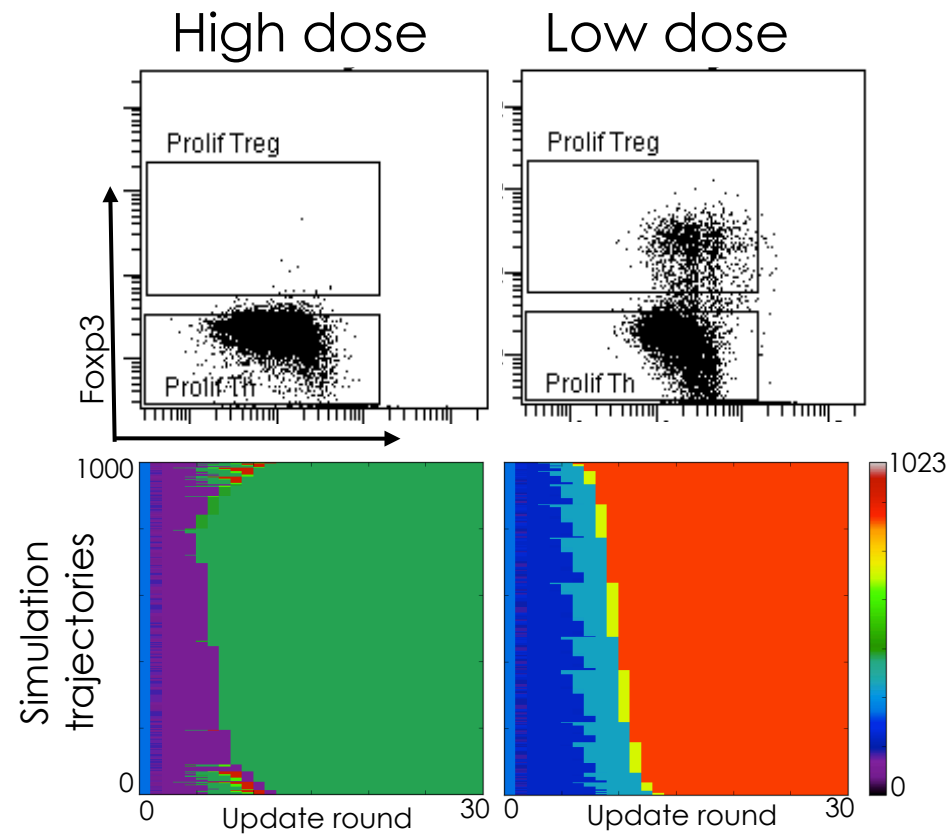
Scenarios 1 and 2: High and Low antigen dose

9



Scenarios 1 and 2: High and Low antigen dose

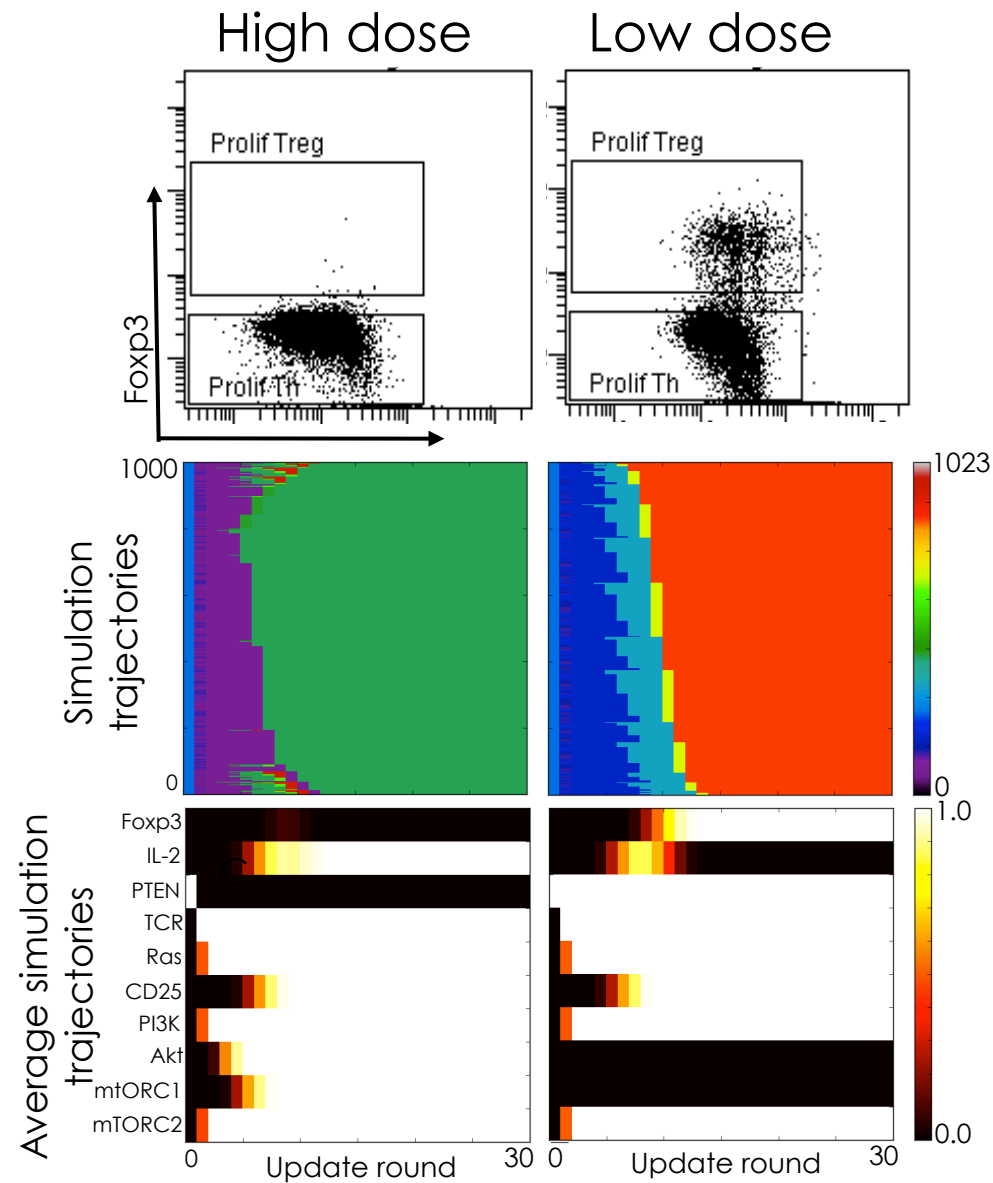
10



Source: N. Miskov-Zivanov et al., in preparation.

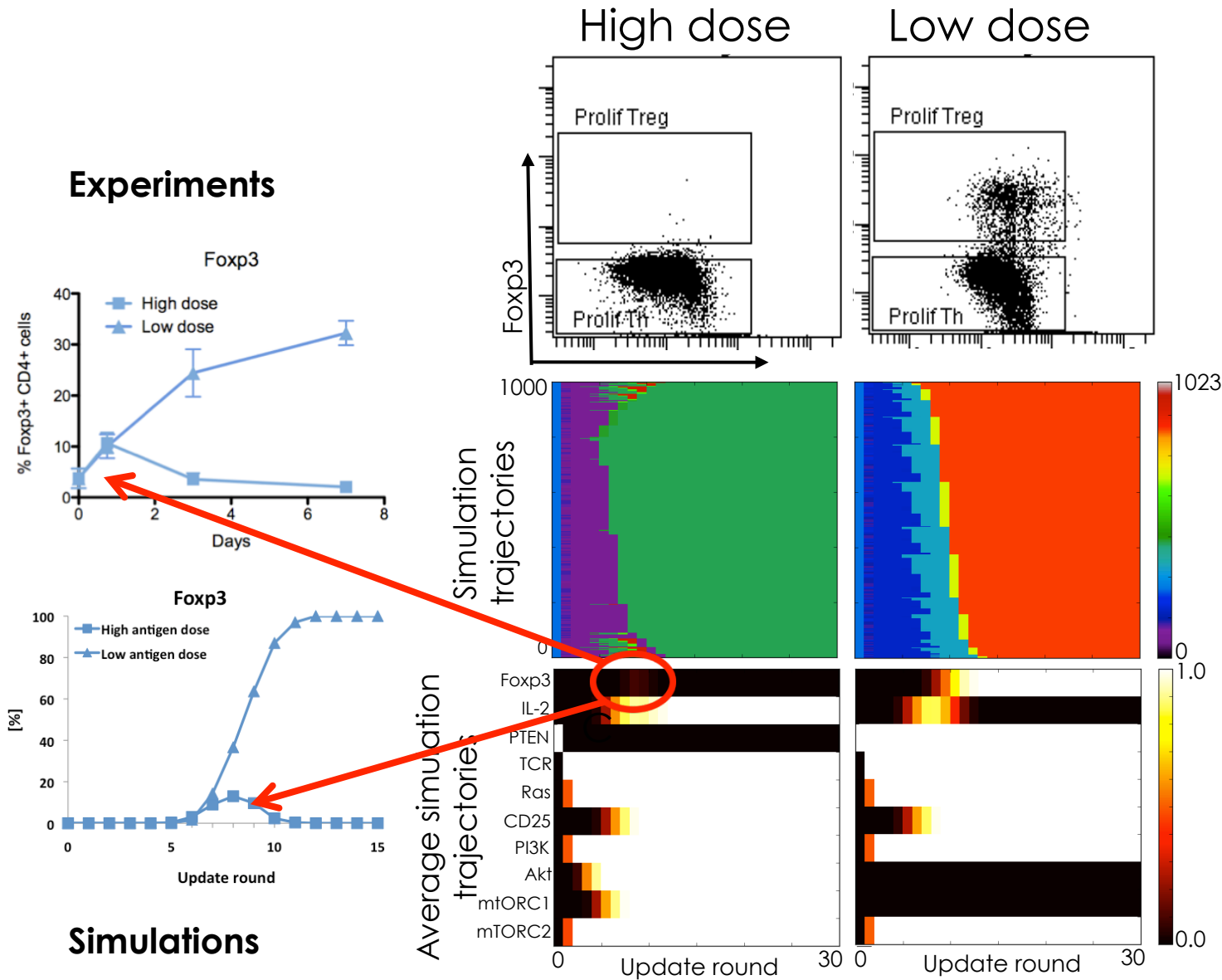
Scenarios 1 and 2: High and Low antigen dose

11



Source: N. Miskov-Zivanov et al., in preparation.

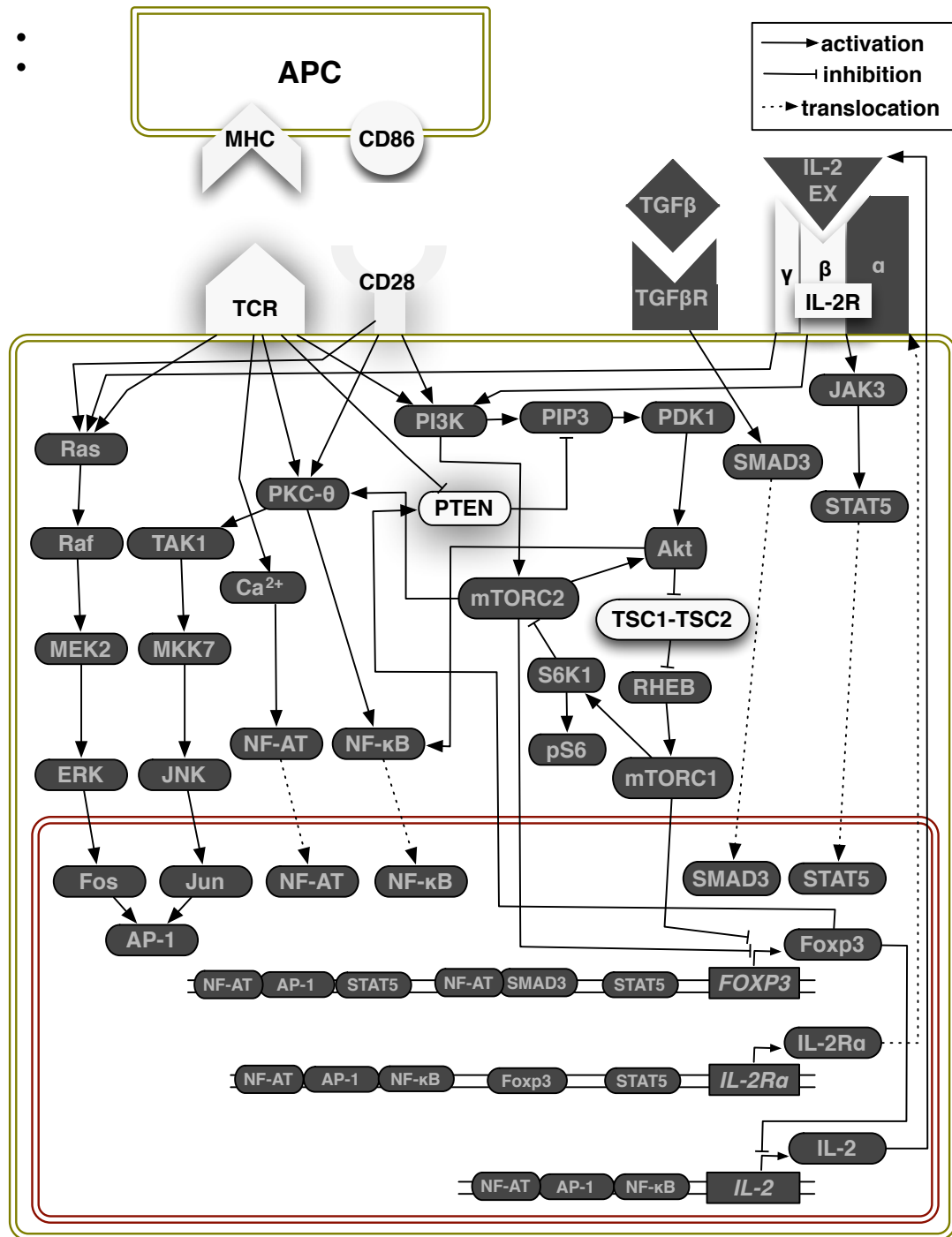
Scenarios 1 and 2: High and Low antigen dose



Source: N. Miskov-Zivanov et al., in preparation.

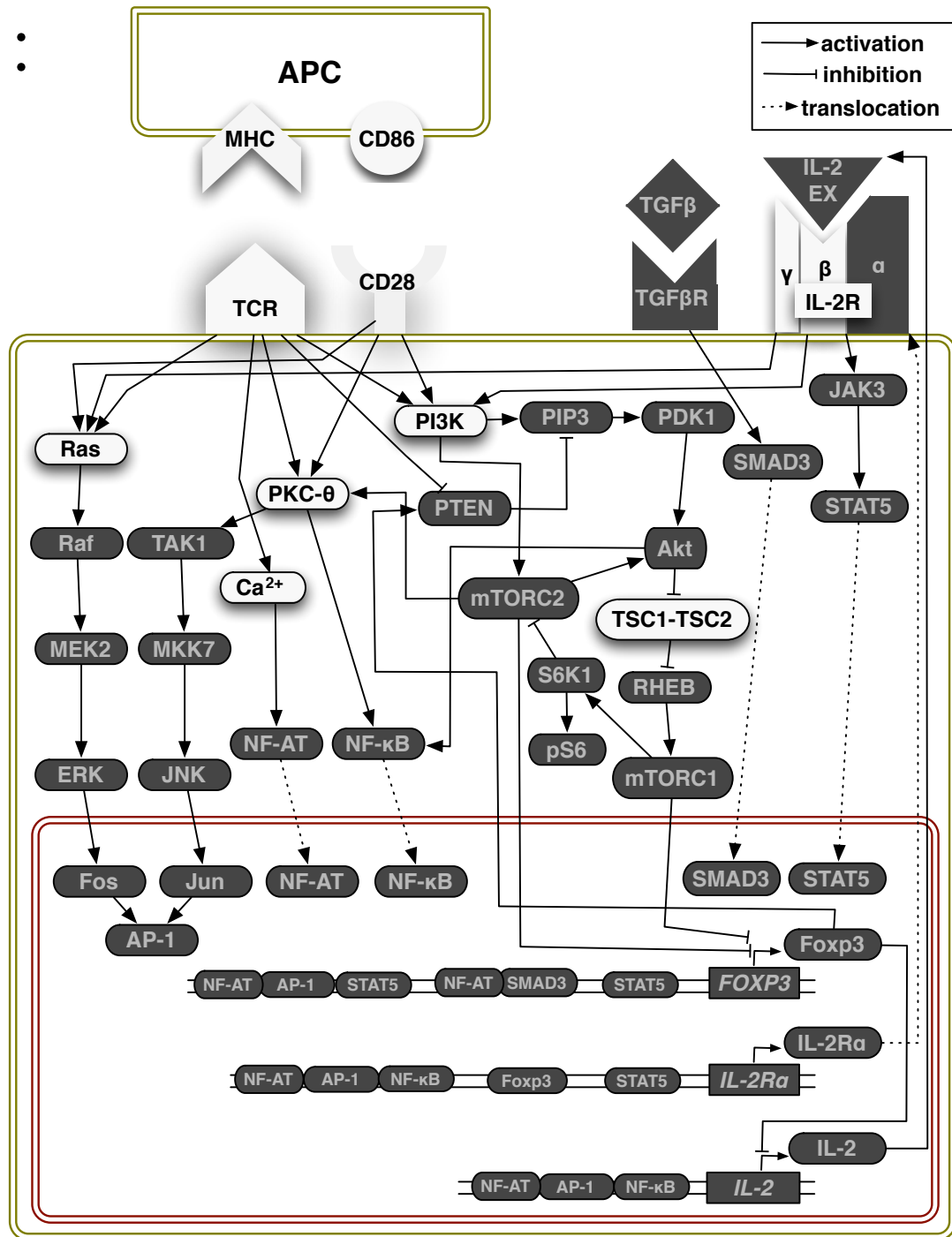
Scenario 1: High antigen dose

value = ON (1)
value = OFF (0)



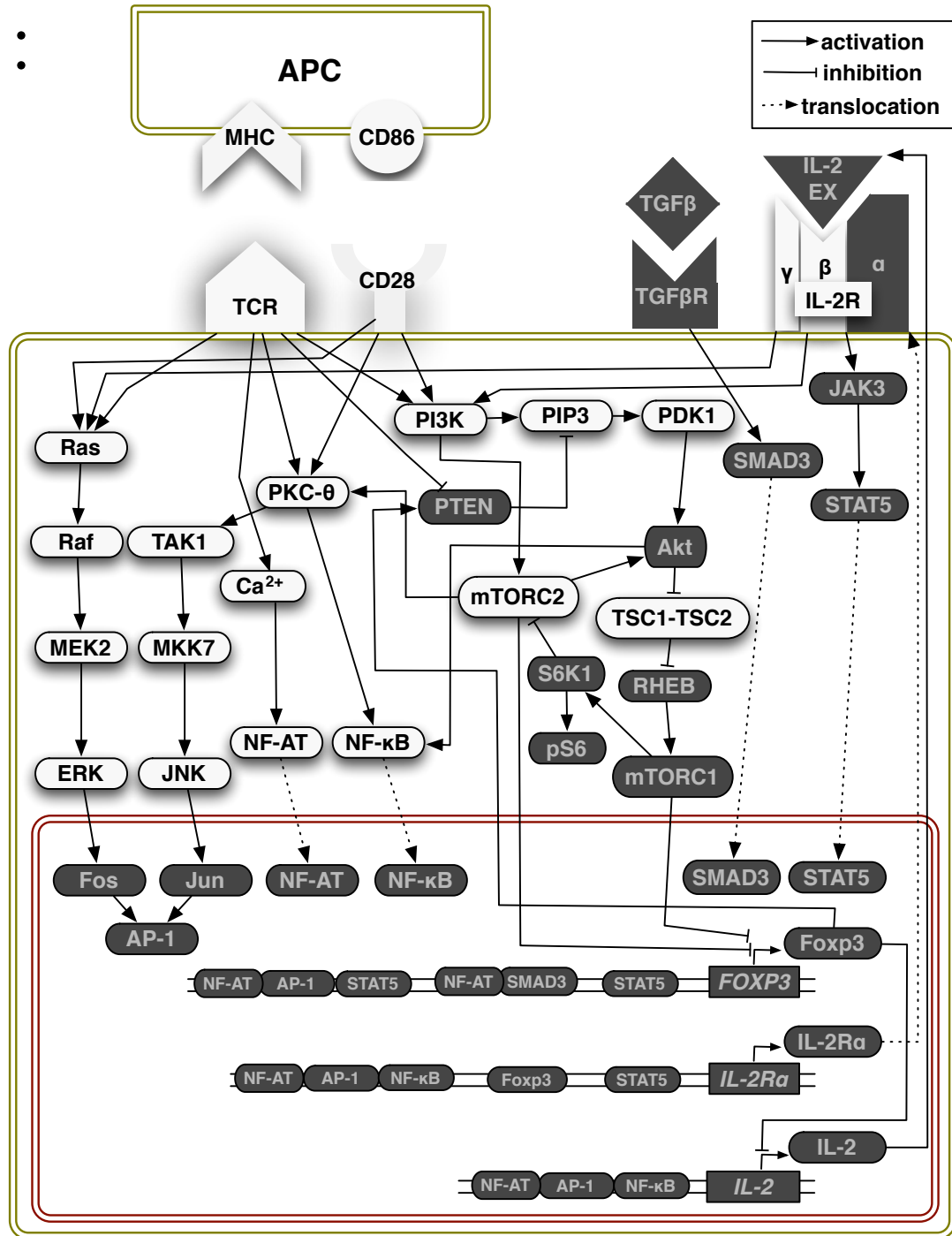
Scenario 1: High antigen dose

value = ON (1)
value = OFF (0)



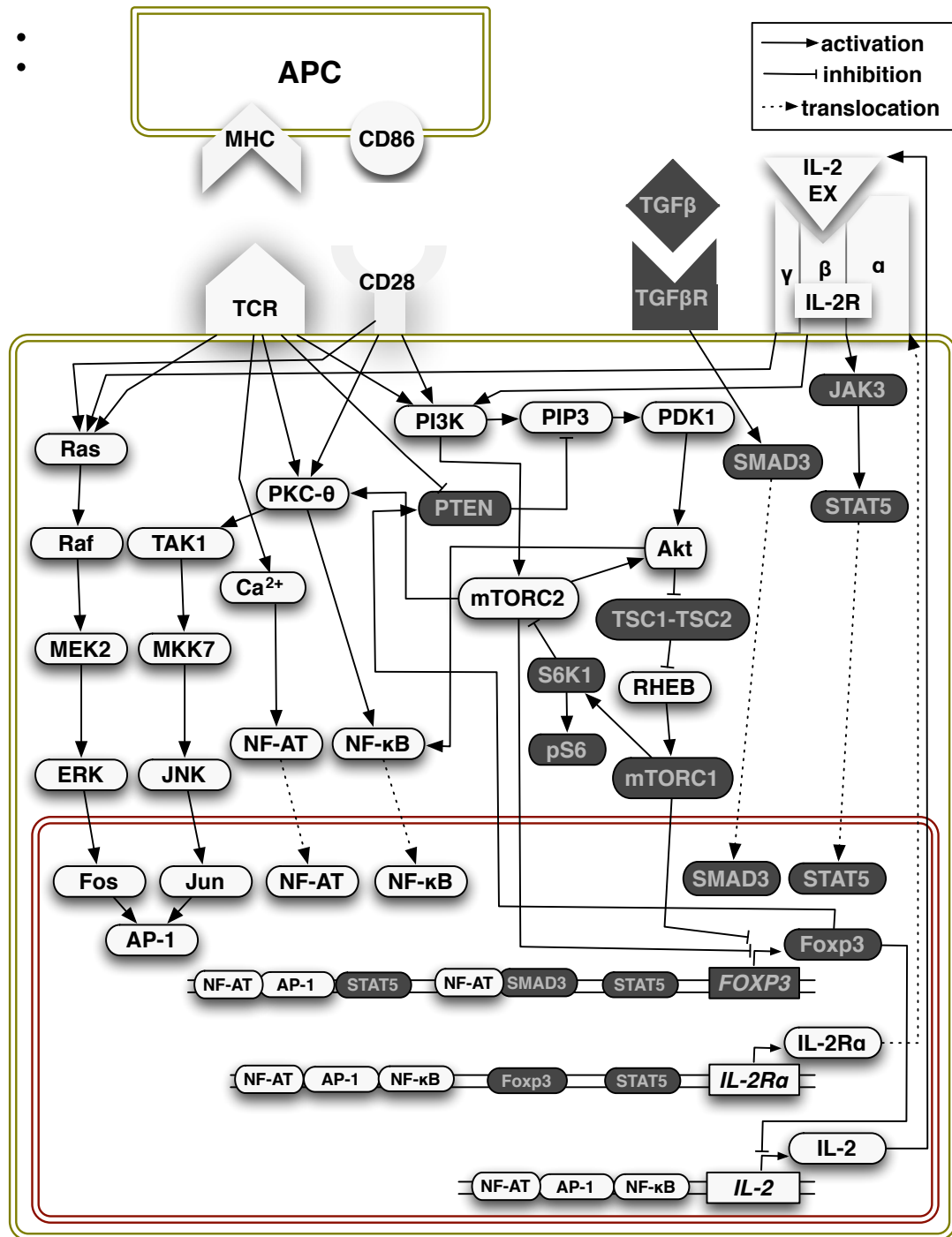
Scenario 1: High antigen dose

value = ON (1)
value = OFF (0)



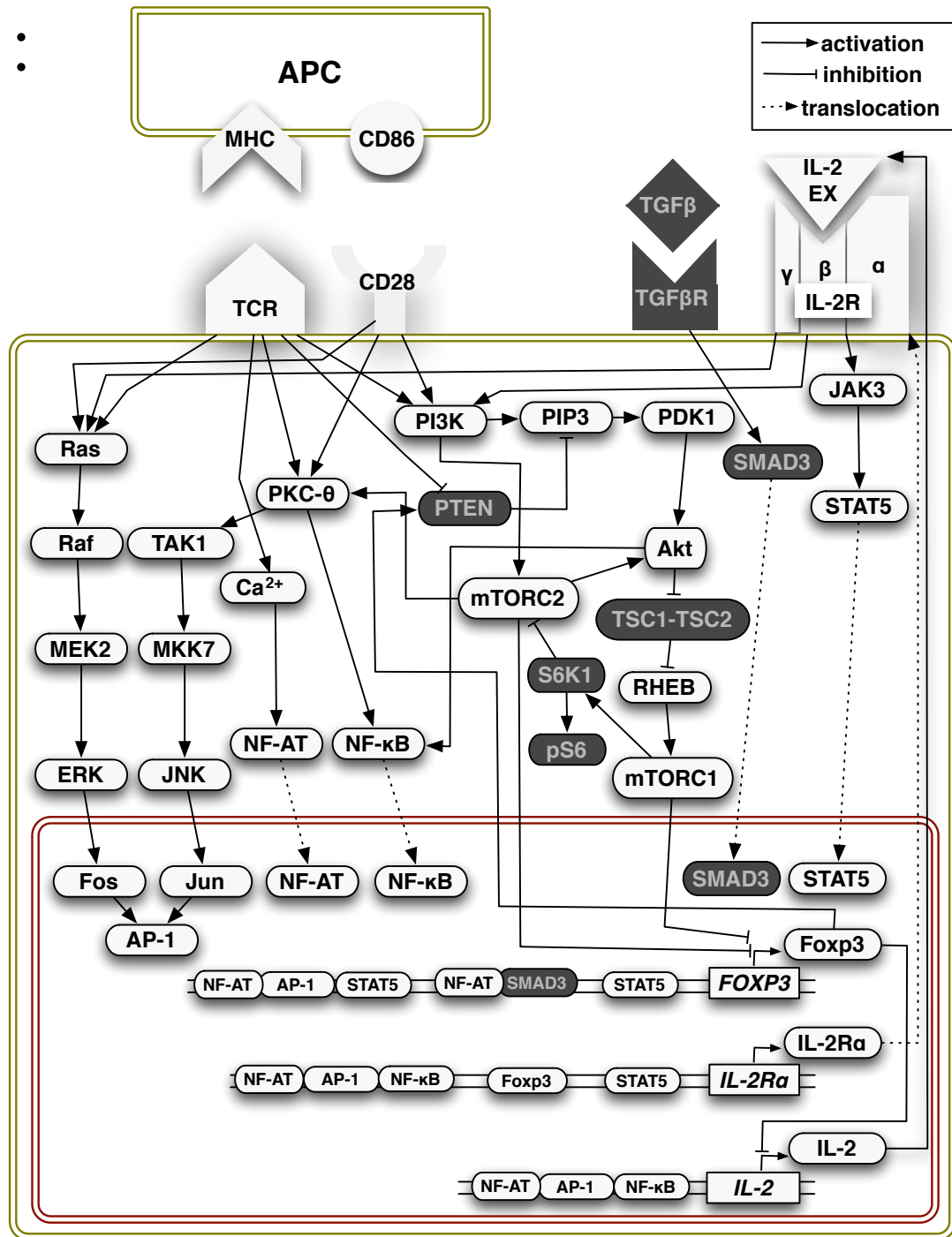
Scenario 1: High antigen dose

value = ON (1)
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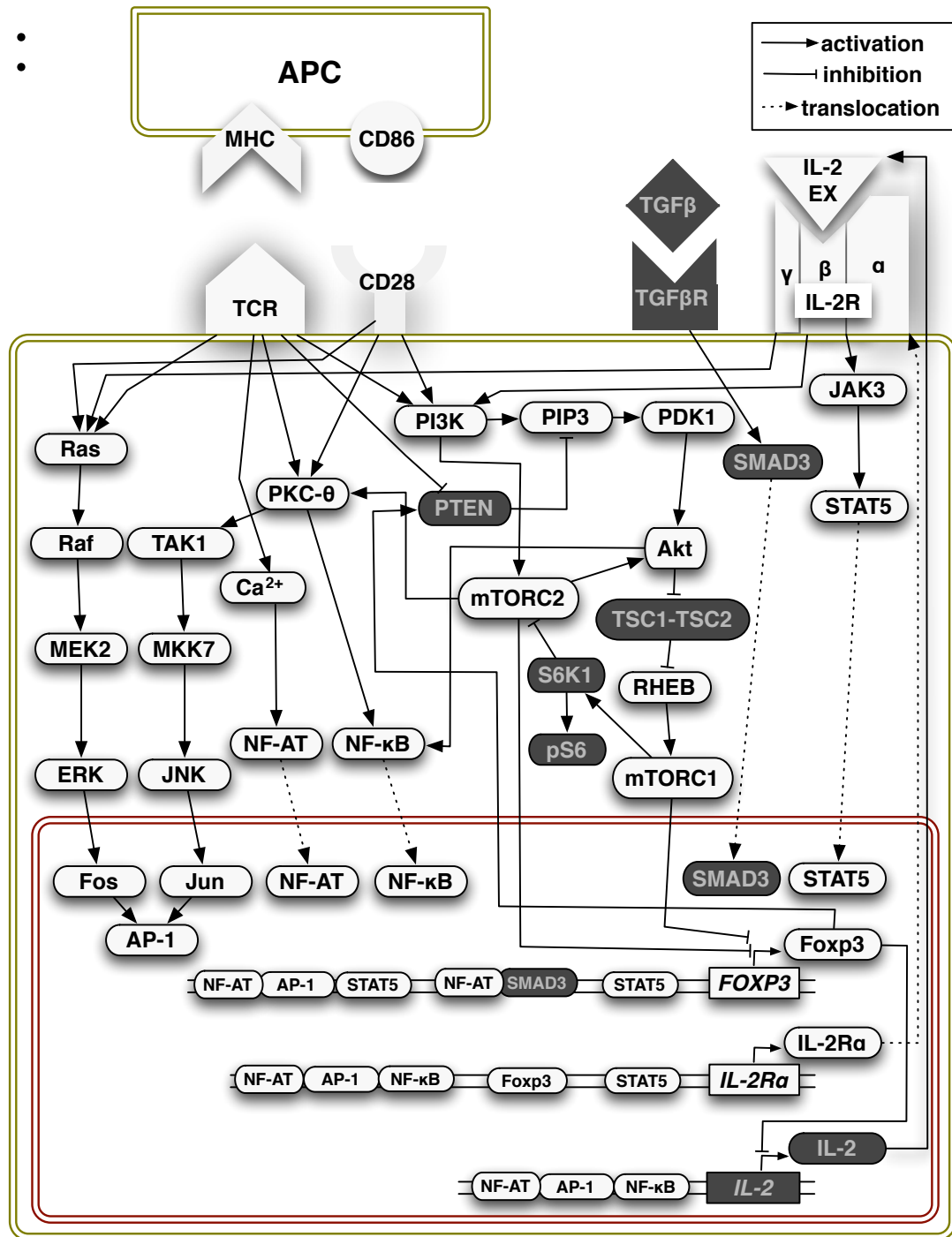
Scenario 1: High antigen dose

value = ON (1)
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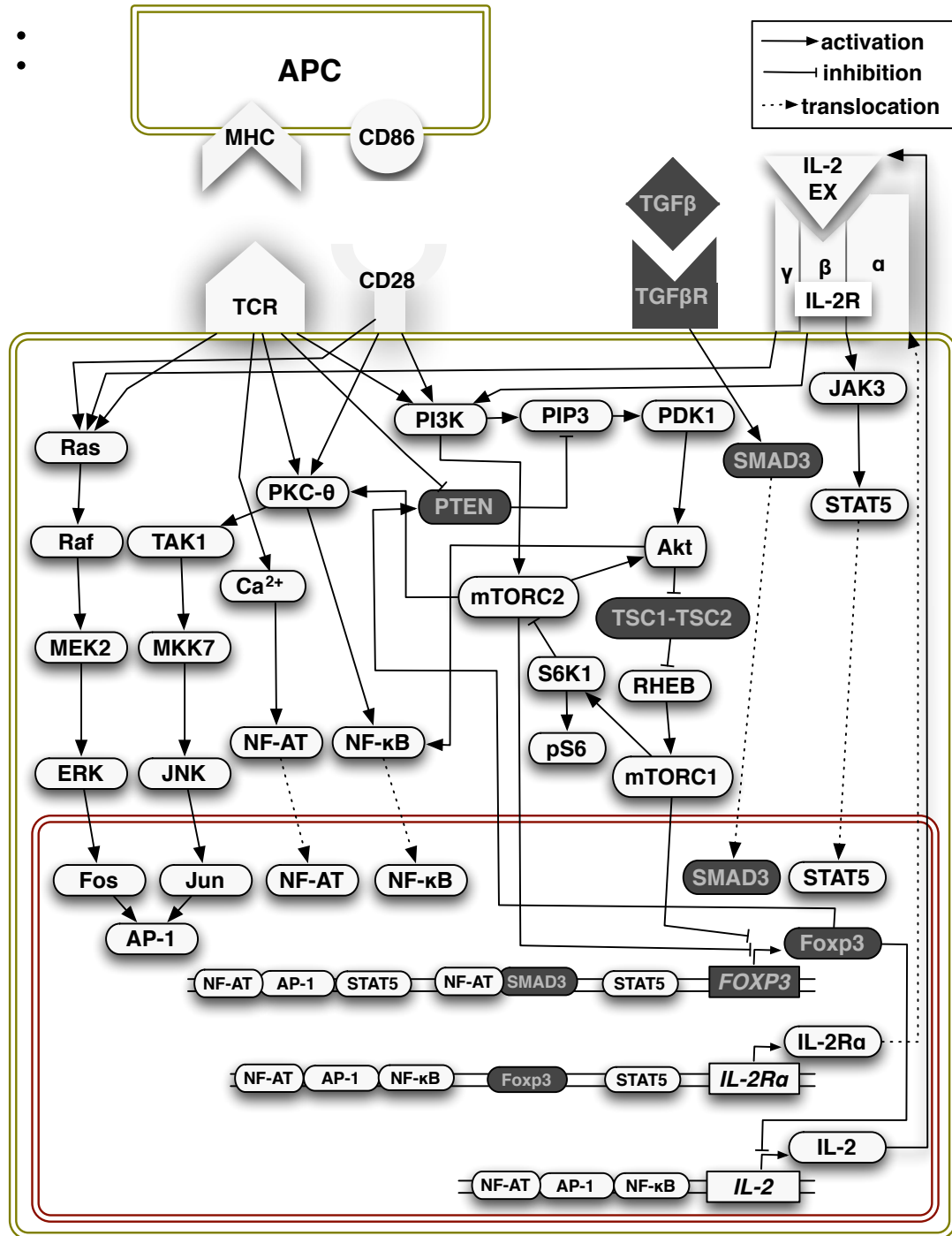
Scenario 1: High antigen dose

value = ON (1)
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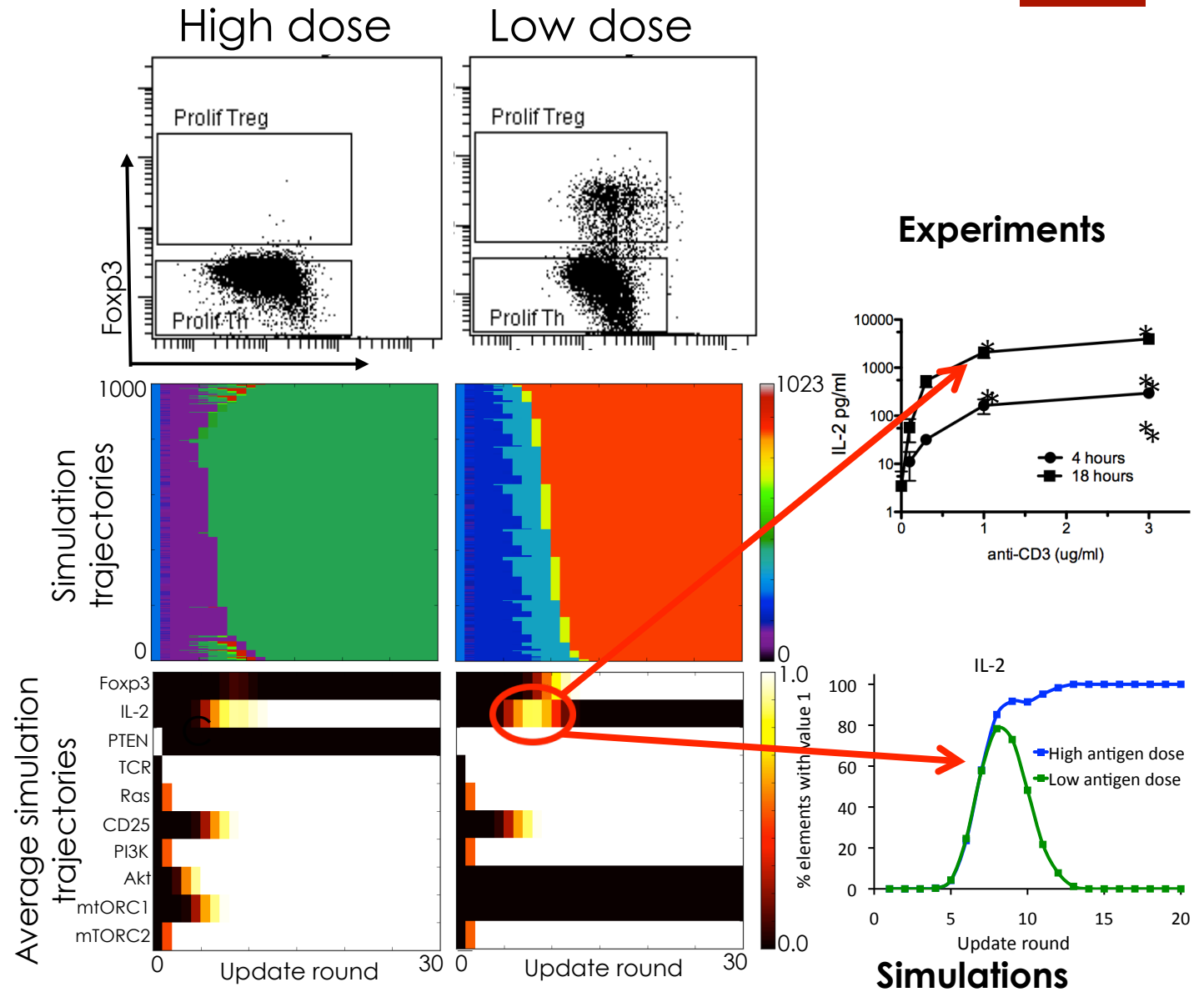


Scenario 1: High antigen dose

value = ON (1)
value = OFF (0)



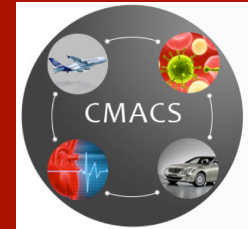
Scenarios 1 and 2: High and Low antigen dose 24



Source: N. Miskov-Zivanov et al., in preparation.

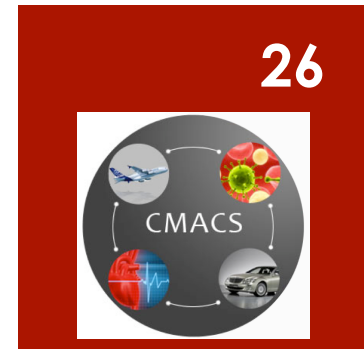
Model does not capture low dose scenario

25

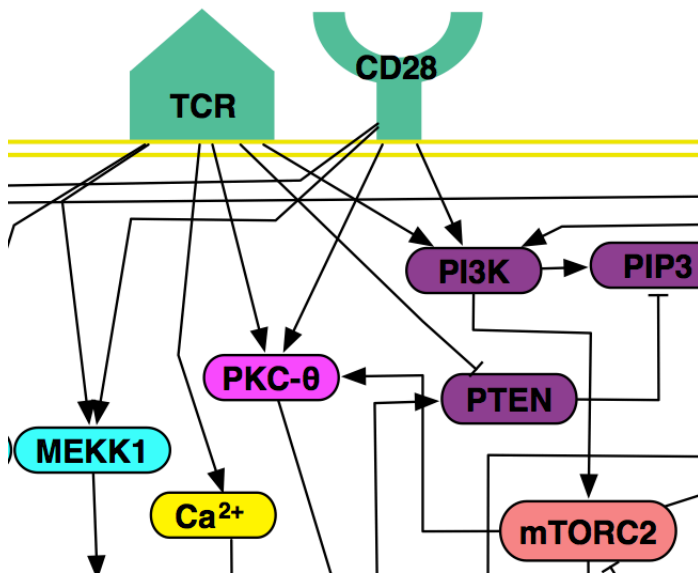


- Model simulations of low antigen dose result in 100% Foxp3+ cells, in experiments no more than 50% Foxp3+ cells

Model does not capture low dose scenario



- Model simulations of low antigen dose result in 100% Foxp3+ cells, in experiments no more than 50% Foxp3+ cells



Low dose antigen is modeled as a change in rules:

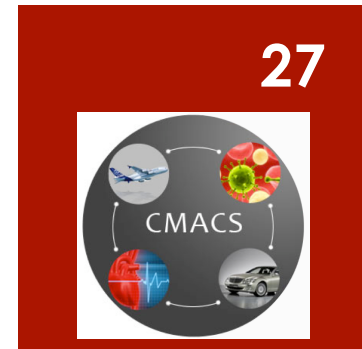
$PKCTHETA^* = TCR_HIGH \text{ or } (TCR_LOW \text{ and } CD28 \text{ and } MTORC2)$

$PI3K_LOW^* = (TCR_LOW \text{ and } CD28) \text{ or } (PI3K_LOW \text{ and } IL2_EX \text{ and } IL2R)$

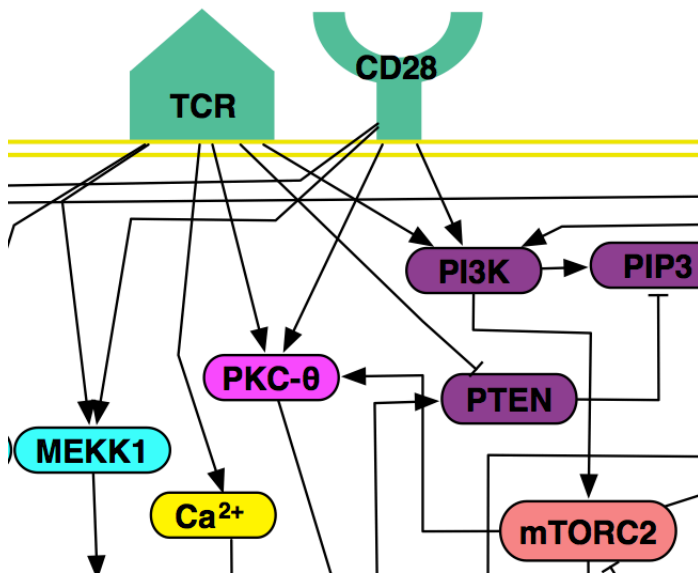
$PI3K_HIGH^* = (TCR_HIGH \text{ and } CD28) \text{ or } (PI3K_HIGH \text{ and } IL2_EX \text{ and } IL2R)$

$PTEN^* = (\text{not } TCR_HIGH \text{ and } PTEN) \text{ or } (\text{not } TCR_HIGH \text{ and } FOXP3)$

Model does not capture low dose scenario



- Model simulations of low antigen dose result in 100% Foxp3+ cells, in experiments no more than 50% Foxp3+ cells



Low dose antigen is modeled as a change in rules:

$PKC_{\theta}^* = TCR_{HIGH} \text{ or } (TCR_{LOW} \text{ and } CD28 \text{ and } mTORC2)$

$PI3K_{LOW}^* = (TCR_{LOW} \text{ and } CD28) \text{ or } (PI3K_{LOW} \text{ and } IL2_{EX} \text{ and } IL2R)$

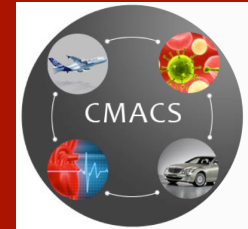
$PI3K_{HIGH}^* = (TCR_{HIGH} \text{ and } CD28) \text{ or } (PI3K_{HIGH} \text{ and } IL2_{EX} \text{ and } IL2R)$

$PTEN^* = (\text{not } TCR_{HIGH} \text{ and } PTEN) \text{ or } (\text{not } TCR_{HIGH} \text{ and } FOXP3)$

 **A more dynamic analysis of TCR signal strength necessary: duration of stimulation**

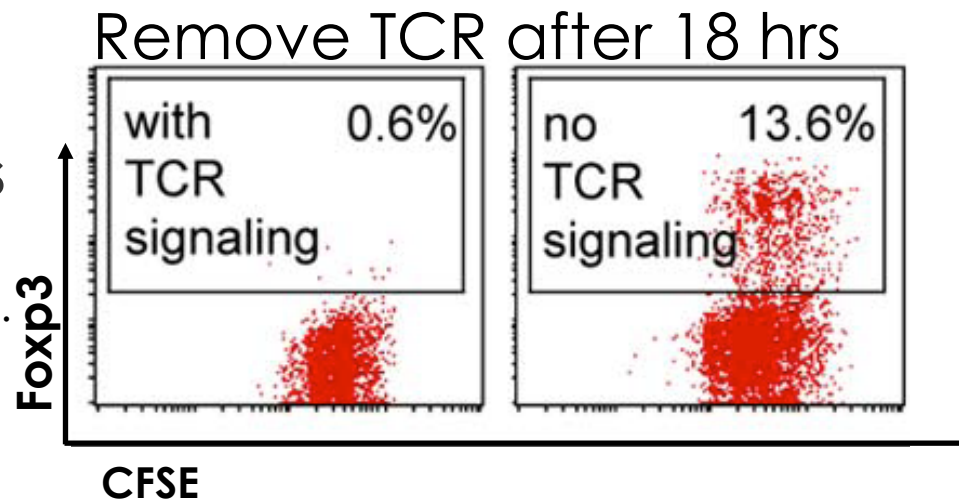
Analysis of duration of stimulation

28



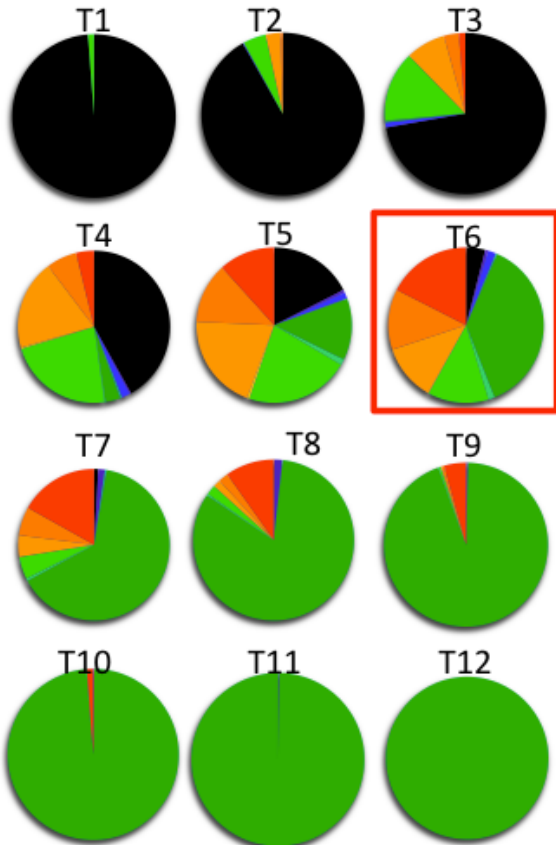
Experiments

Source: Sauer *et al.*,
PNAS 105:7797, 2008.

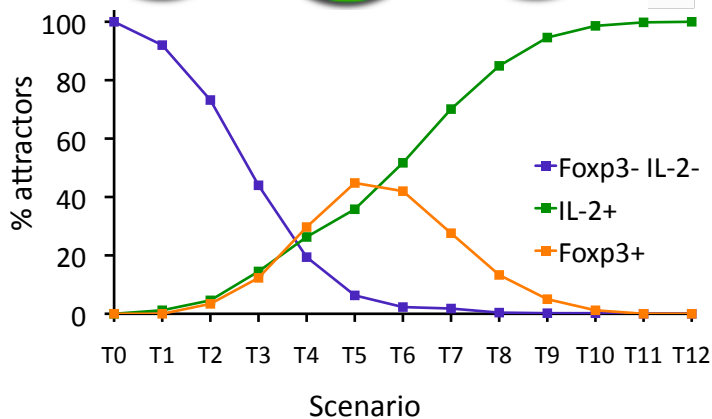


Scenario 3:

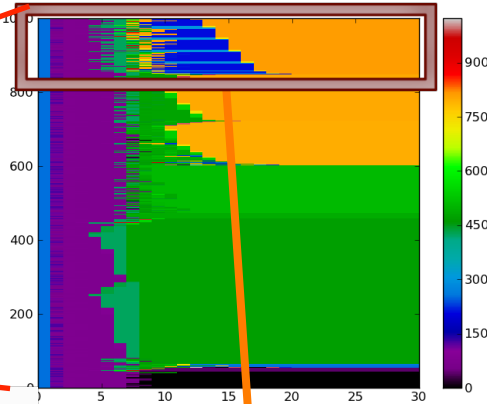
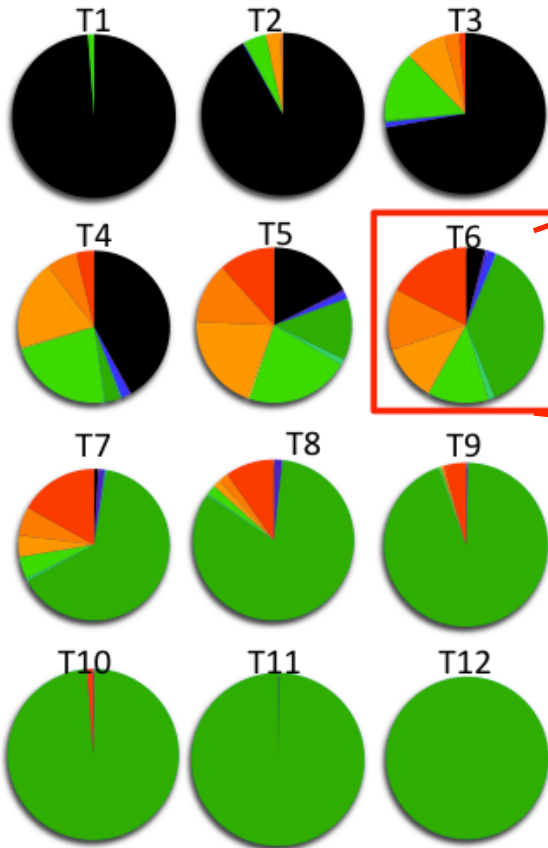
Antigen removal at rounds 1-12 (T1-T12)



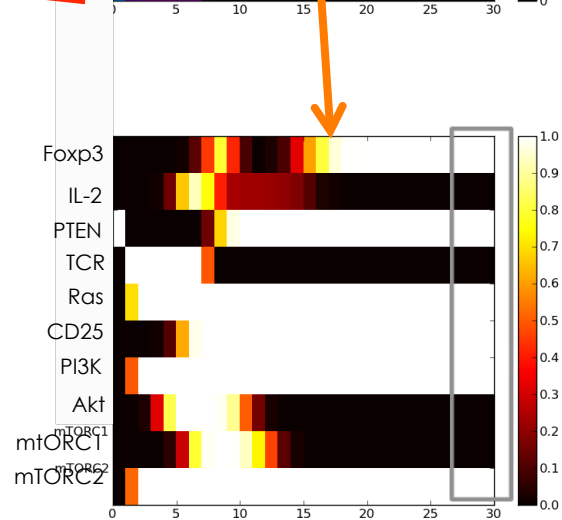
Attractors	T6											No removal	
	A1	A2	A3	A4	A5	A6	A7	A8	A9	A10	A11	HD	LD
Foxp3	█	█	█	█	█	█	█	█	█	█	█	█	█
IL-2	█	█	█	█	█	█	█	█	█	█	█	█	█
PTEN	█	█	█	█	█	█	█	█	█	█	█	█	█
TCR	█	█	█	█	█	█	█	█	█	█	█	█	█
Ras	█	█	█	█	█	█	█	█	█	█	█	█	█
CD25	█	█	█	█	█	█	█	█	█	█	█	█	█
PI3K	█	█	█	█	█	█	█	█	█	█	█	█	█
Akt	█	█	█	█	█	█	█	█	█	█	█	█	█
mTORC1	█	█	█	█	█	█	█	█	█	█	█	█	█
mTORC2	█	█	█	█	█	█	█	█	█	█	█	█	█
Attractor size	40	6	17	3	374	13	127	1	118	126	175	1000	1000



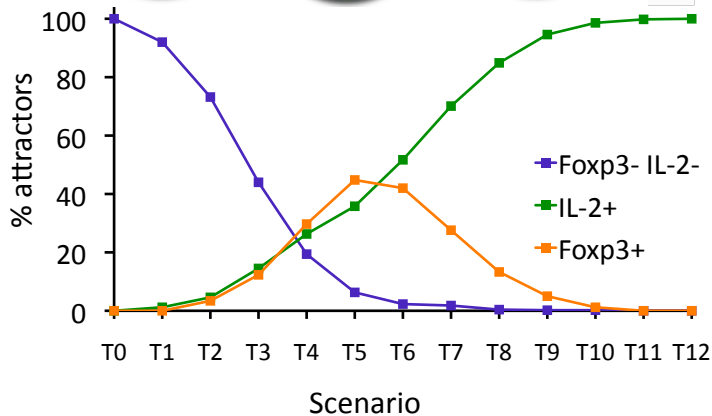
Scenario 3: Antigen removal at rounds 1-12 (T1-T12)



1000 simulation trajectories



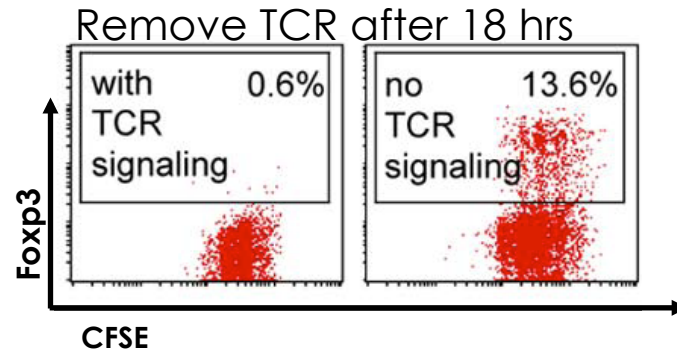
Average trajectories for attractor 1010111001



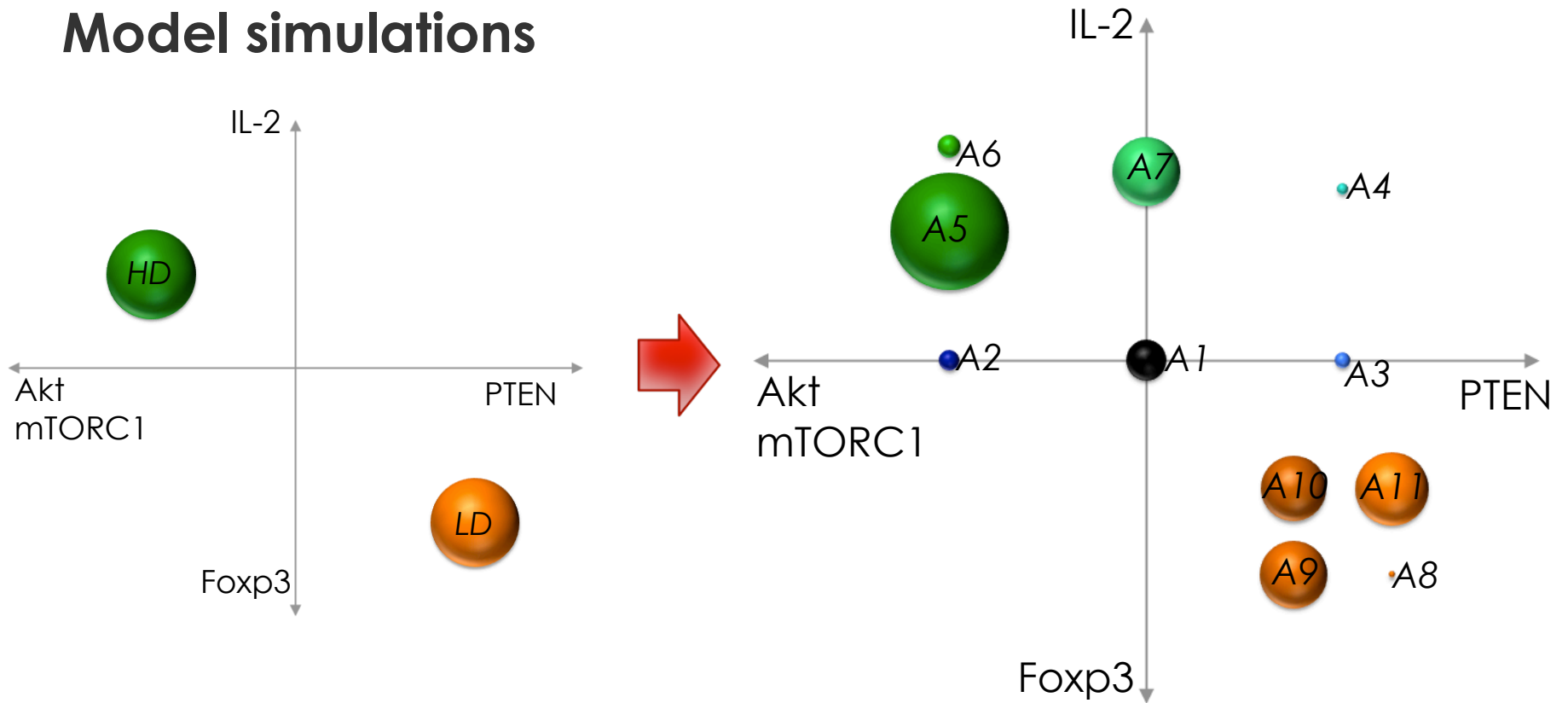
Scenario 3: Antigen removal at round 6 (T6)

Experiments

Source: Sauer et al., PNAS 105:7797, 2008.



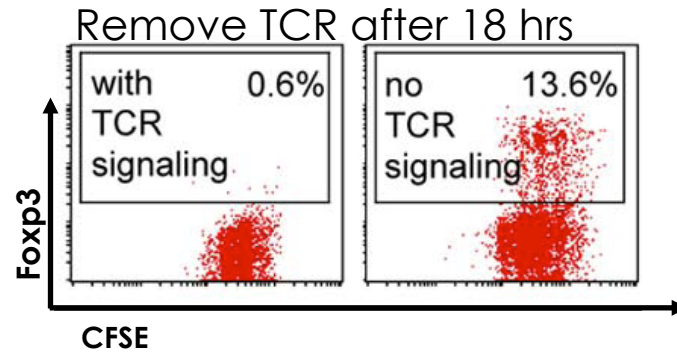
Model simulations



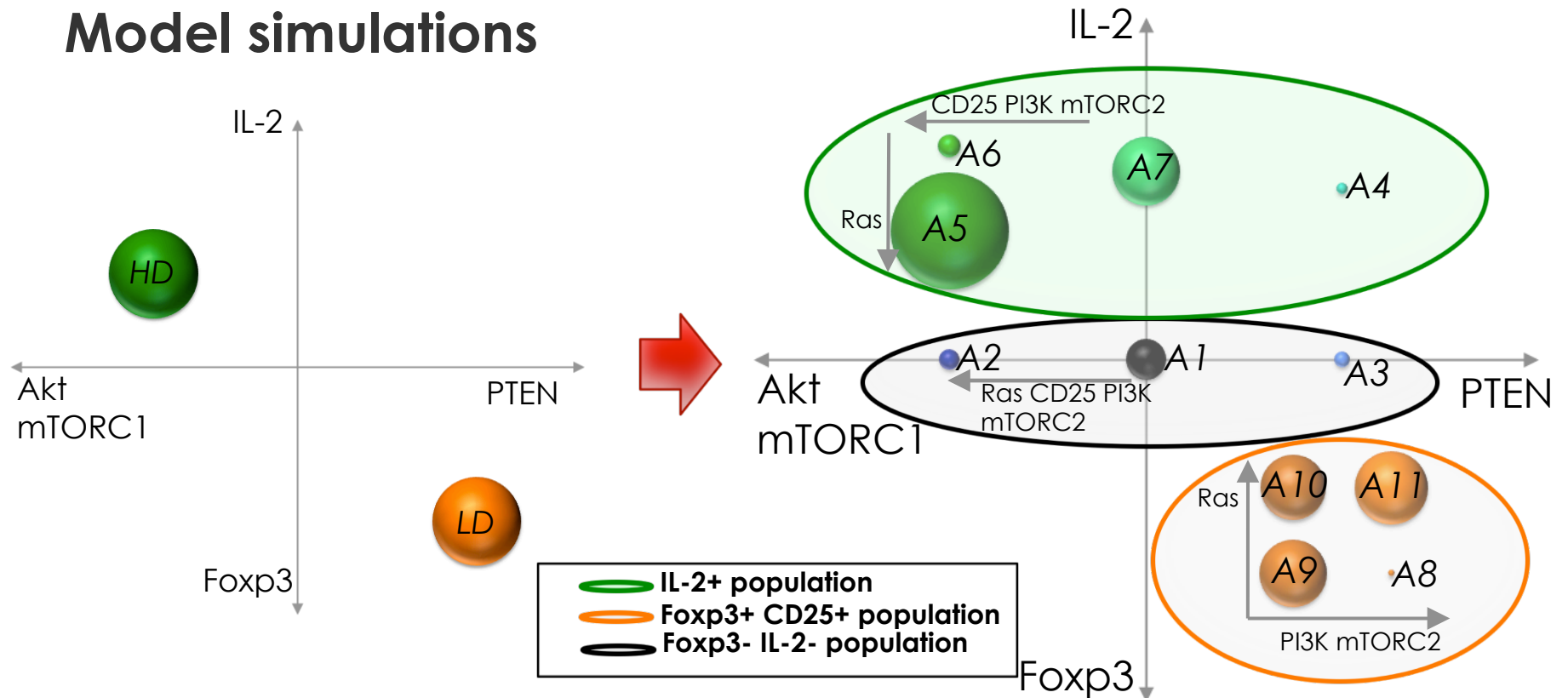
Scenario 3: Antigen removal at round 6 (T6)

Experiments

Source: Sauer et al., PNAS 105:7797, 2008.

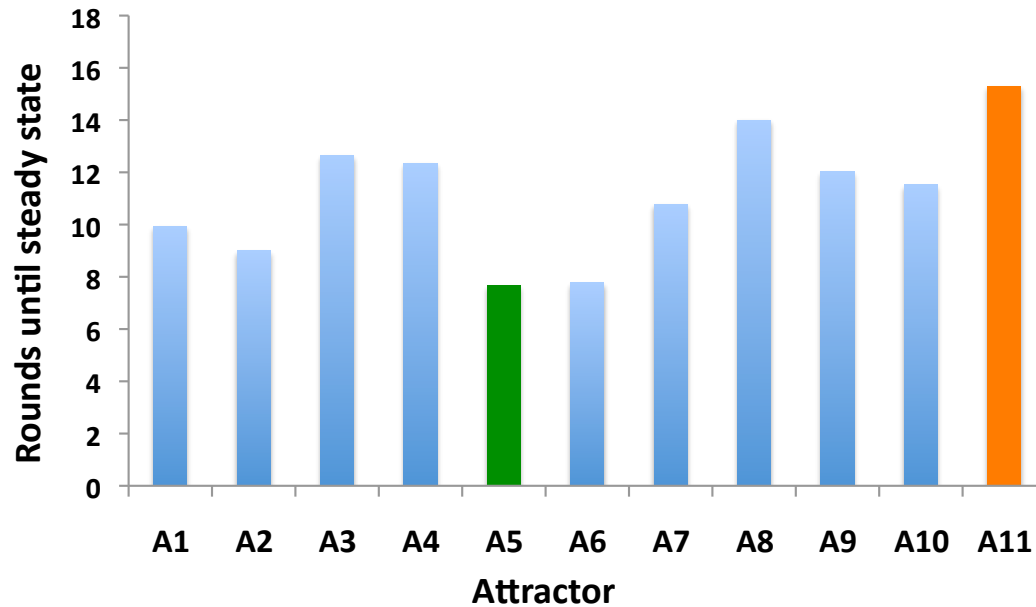


Model simulations

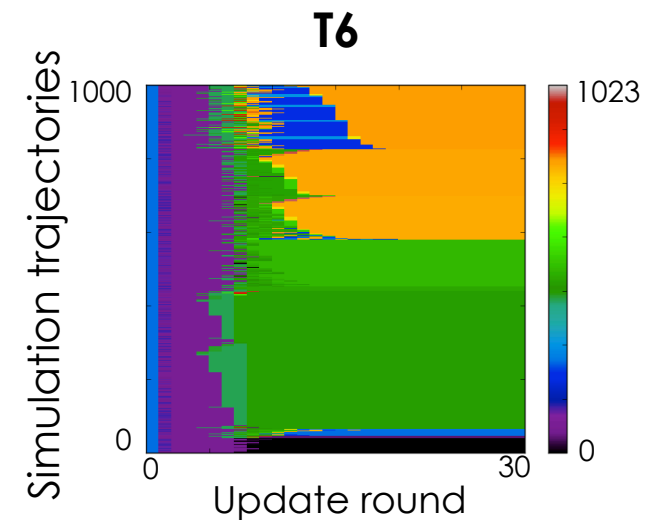
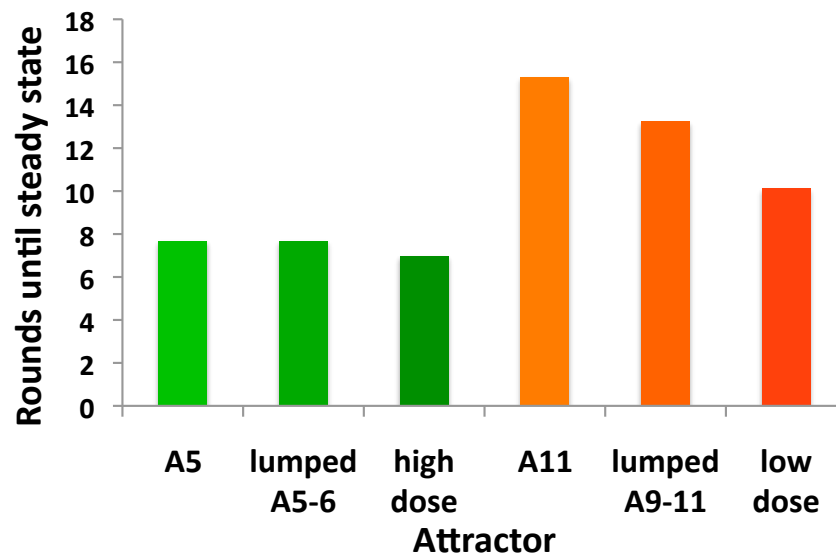


Time to reach steady state (T6, HD, LD)

33

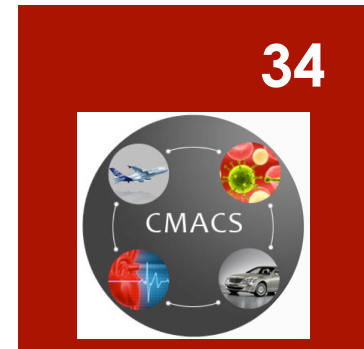


■ Treg cells take longer to differentiate than Th cells



Model checking

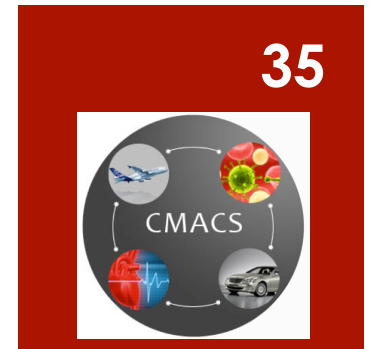
- SPIN provides yes/no answers



SPIN					
High dose			Low dose		
Specification	LTL Formula	Verified	Specification	LTL Formula	Verified
Does Foxp3 become false forever?	$!(\langle \rangle [] !\text{foxp3})$	Yes	Does Foxp3 become true forever?	$!(\langle \rangle [] \text{foxp3})$	Yes
Does ps6 become true forever?	$!(\langle \rangle [] \text{ps6})$	Yes	Does pS6 stay false forever?	$!([\] !\text{ps6})$	Yes
Does PIP3 become true forever?	$!(\langle \rangle [] \text{pip3})$	Yes	Does IL-2 become true but eventually become false forever?	$!(\langle \rangle i12 \ \&\& \ \langle \rangle [] !i12)$	Yes
Does Akt become true forever?	$!(\langle \rangle [] \text{akt})$	Yes	Is PTEN always true?	$!([\] \text{pten})$	Yes
Does mtTORC1 become true forever?	$!(\langle \rangle [] \text{mtorc1})$	Yes	Does mTORC become true forever?	$!(\langle \rangle [] \text{mtorc})$	Yes
Does S6K1 become true forever?	$!(\langle \rangle [] \text{s6k1})$	Yes	Does CD25 become true forever?	$!(\langle \rangle [] \text{cd25})$	Yes
Does STAT5 become true forever?	$!(\langle \rangle [] \text{stat5})$	Yes	Does STAT5 become true forever?	$!(\langle \rangle [] \text{stat5})$	Yes
Does CD25 become true forever?	$!(\langle \rangle [] \text{cd25})$	Yes	High dose + TGFβ		
Does IL-2 become true forever?	$!(\langle \rangle [] i12)$	Yes	Does IL-2 become true forever?	$!(\langle \rangle [] i12)$	Yes
Does PTEN become true forever?	$!(\langle \rangle [] \text{pten})$	Yes	Does PTEN become false forever?	$!(\langle \rangle [] !\text{pten})$	Yes

Probabilistic model checking

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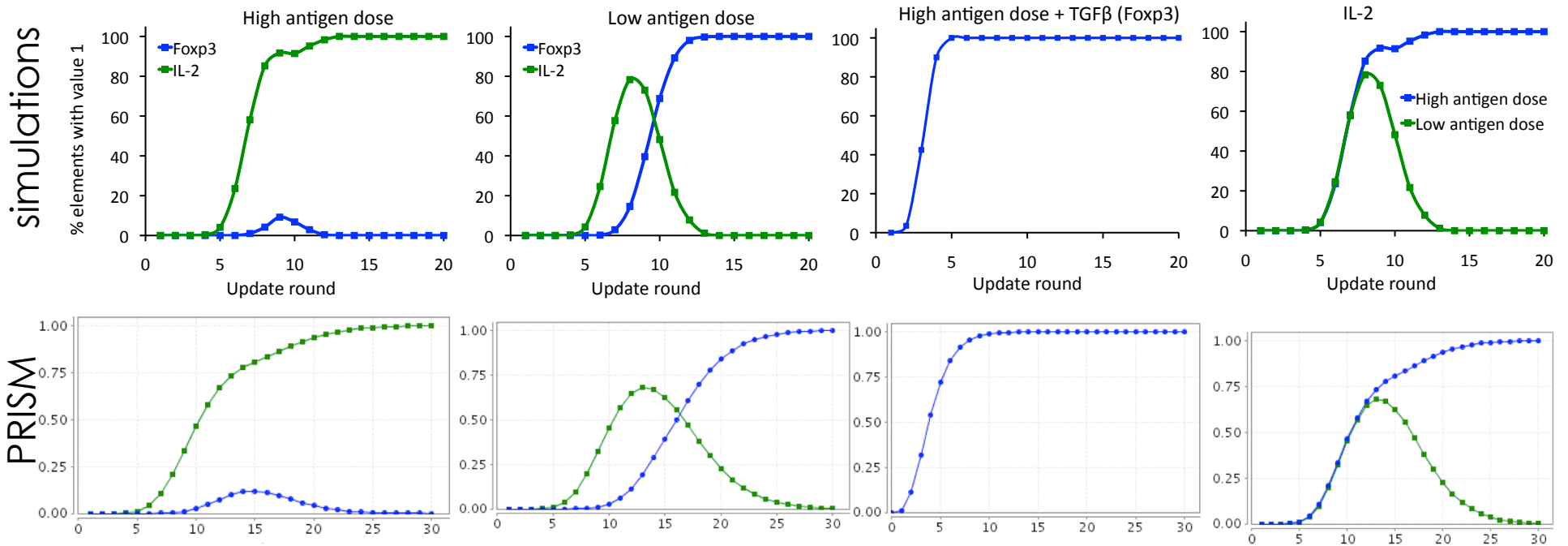
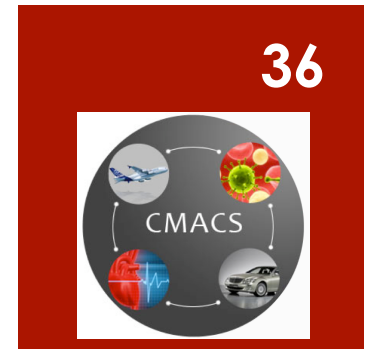


- PRISM allows for analyzing transient behavior

PRISM					
High dose			Low dose		
Specification	LTL Formula	Prob.	Specification	LTL Formula	Prob.
Does Foxp3 eventually become false forever?	$P=? [FG! \text{foxp3}]$	1	Does pS6 stay true?	$P=? [G \text{ps6}]$	0
What is the probability that Foxp3 never becomes true?	$P=? [G! \text{foxp3}]$	0.765098	Does mTORC2 ever get to true?	$P=? [F \text{mtorc2}]$	1
Does ps6 become true forever?	$P=? [F G \text{ps6}]$	1	Does PTEN ever get to false?	$P=? [F !\text{pten}]$	0
Are IL-2 and PTEN ever both true?	$P=? [F ((\text{pten}) \& (\text{il2}))]$	0.000468	High dose + TGFβ		
What is the probability that IL-2 is false before Foxp3 becomes true	$P=? [(!\text{il2})U(\text{foxp3})]$	0	Does IL-2 become true?	$P=? [F \text{il2}]$	0.019584
PRISM – behavior over time					
What is the behavior of Foxp3 over time?	$P=? [G[t,t] \text{foxp3}]$	Figure	What is the behavior of IL-2 over time?	$P=? [G[t,t] \text{il2}]$	Figure

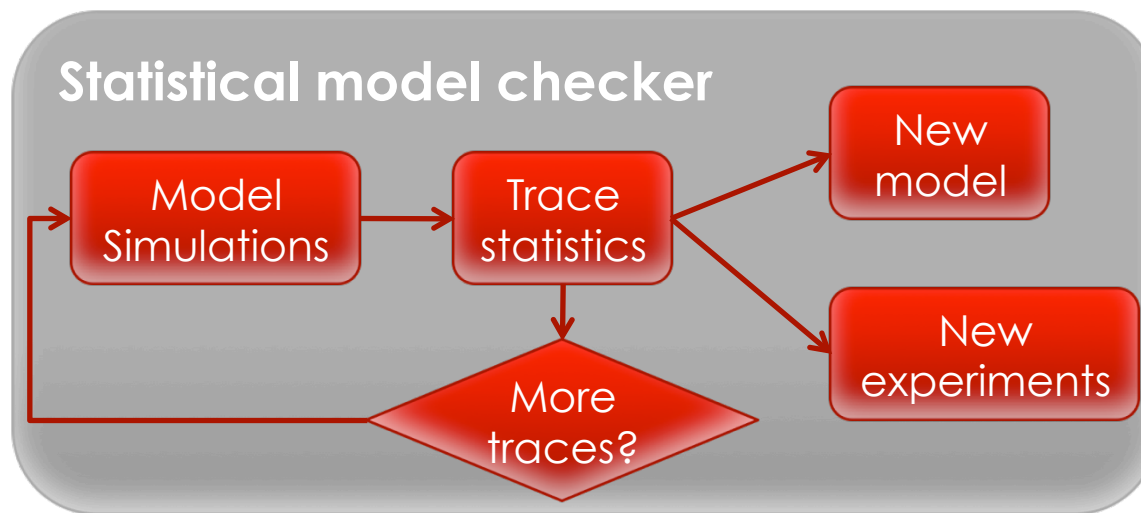
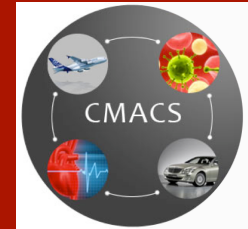
Probabilistic model checking

- PRISM allows for analyzing transient behavior



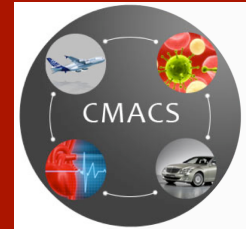
Statistical model checking

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Statistical model checking

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- Low antigen dose scenario:

1. Does IL-2 always go to 1?

Property: $F[20] (IL2 == 1)$

Test: BEST 0.001 0.999

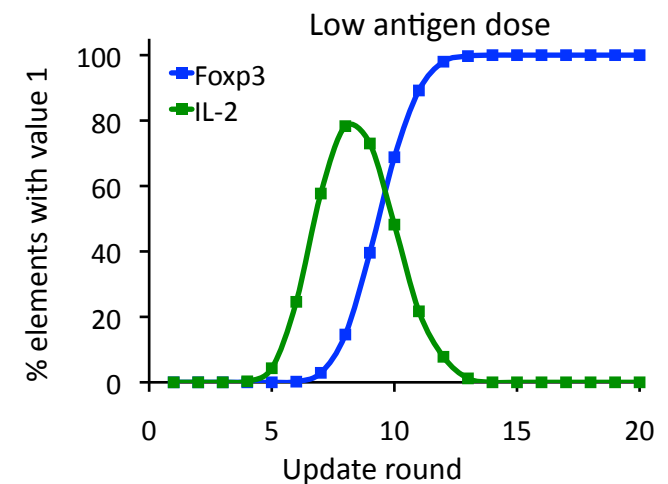
Result: estimated probability close to 1

2. Probability that IL-2 stays at 0 before Foxp3 becomes 1?

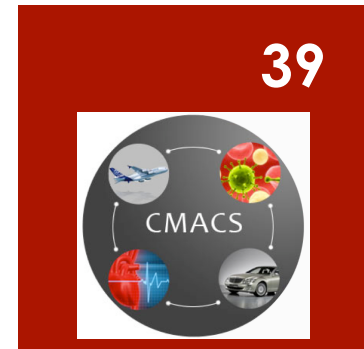
Property: $(IL2 == 0) U[15] (FOXP3 == 1)$

Test: BEST 0.0001 0.999

Result: estimated probability = 0.00147
rare event

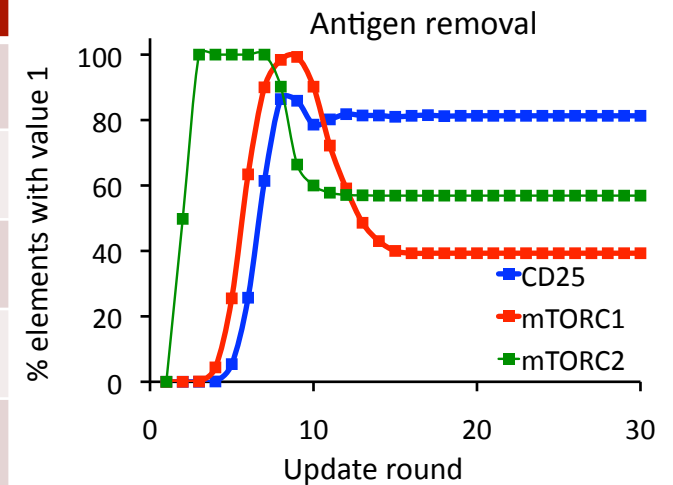


Statistical model checking



- High antigen dose + antigen removal scenario:
 - Studies of relative timing on mTOR vs. CD25/STAT5 pathway

	Property	Probability estimate and sample size	Elapsed time [s]
1	$G^7 \sim (\text{MTORC1} = 1 \ \& \ \text{MTORC2} = 1)$	estimate = 0.0188048 samples = 200,160	1,946
2	$F^7 (\text{MTORC1} = 1 \ \& \ \text{MTORC2} = 1)$	estimate = 0.980884 samples = 2,352	23
3	$F^{10} (\text{MTORC1} = 1 \ \& \ \text{MTORC2} = 1 \ \& \ \text{CD25} = 0 \ \& \ (F^{18} (\text{CD25} = 1)))$	estimate = 0.60104 samples = 25,968	253
4	$F^{28} (\text{MTORC1} == 1 \ \& \ \text{MTORC2} == 1 \ \& \ \text{CD25} == 0 \ \& \ (F^1 (\text{CD25} == 1)))$	estimate = 0.592195 samples = 26,160	254
5	$F^{10} (\text{MTORC1} = 1 \ \& \ \text{MTORC2} = 1 \ \& \ \text{CD25} = 0 \ \& \ (F^1 (G^{17} (\text{CD25} = 1))))$	estimate = 0.39669 samples = 25,920	254



Conclusion

- **Logical modeling approach allows development of comprehensive models of cell fate**

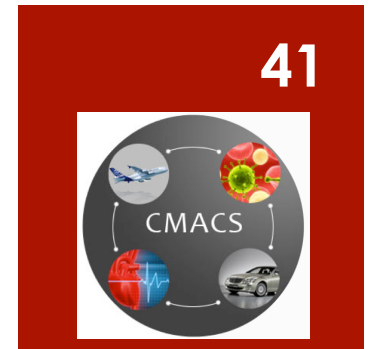
- Model of peripheral T cell differentiation recapitulates a wide range of experimental observations
- Circuit analysis reveals key elements of the mechanism for Foxp3 expression
- Timing is critical for Treg differentiation:
 - Treg cells take longer to differentiate than Th cells
 - Race between Foxp3 activating and inhibiting pathways
 - Feedback between Foxp3 and PTEN



Conclusion

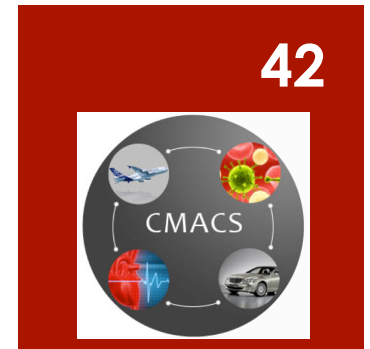
■ Model checking

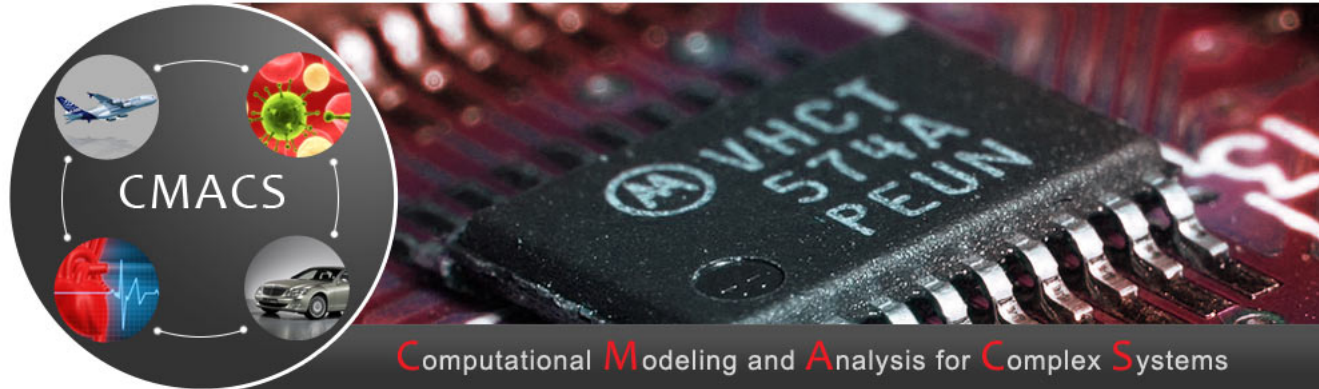
- Allows for more efficient studies of the model
- Probabilistic model checking:
 - Provides transient results that match simulations
- Statistical model checking:
 - Further analysis of of transient behavior
 - Provides insights into timing relationships between elements



Next steps

- Analyze different removal scenarios using model checking
- Expansion of the model (keep up with fast pace of developments in the field)
- Develop a model for several cell types
- Develop population models that embed intracellular circuitry





Thank you!